Detection of *Giardia* cysts in sewage and estimations of giardiasis prevalence among inhabitants in Hyogo Prefecture, Japan

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**Abstract:** We recovered *Giardia* cysts from sewage and estimated the prevalence of giardiasis among inhabitants in Hyogo Prefecture, western Japan. Two 200-ml sewage samples were collected once a month from two different sewage plants (A and B) during a one-year period. After pretreating the samples by filtration and centrifugation in the presence of alcohol and diethyl ether, an immunomagnetic separation method was applied to isolate *Giardia* cysts. The mean number of cysts recovered from plants A and B were 880/L and 2,000/L, respectively. Circadian rhythms in the number of cysts detected on each of two examination days were observed with a peak at 10:00 a.m., and seasonal variations in cyst numbers were also observed with two peaks in May and December. By using the data obtained from this survey, we estimated that the prevalence of *Giardia* infection in inhabitants in the areas serviced by plants A and B were 0.1-1.1 and 0.1-1.3%, respectively.

**Key Words:** *Giardia intestinalis*, sewage, detection method, prevalence

**INTRODUCTION**

The flagellate protozoan parasite *Giardia intestinalis* is widely distributed throughout the world with a high prevalence in developing countries in the tropics and subtropics. *G. intestinalis* infection is acquired by digesting cysts excreted from infected humans and animals [1]. Approximately 20 to 30% of inhabitants in developing countries in Asia and Africa are infected with this parasite. In developed countries with improved hygiene conditions, meanwhile, *Giardia* infection also constitutes a public health concern [2]. In developed countries, the parasite is brought back from abroad by infected travelers [3]. Many outbreaks of giardiasis due to ingestion of inadequately treated water have also occurred in developed countries; five of these occurred in the United States in 1993 and 1994 [4]. Although a direct smear method is frequently used to detect *Giardia* cysts in stool samples, it does not consistently reflect the accurate prevalence of giardiasis in a given survey area. For instance, 15 to 20% of patients are reportedly diagnosed as false negative [5], despite the use of appropriate fecal examinations. The need for examination of three consecutive stool samples has therefore been indicated for accurate diagnosis [6,7].

The reported prevalence of giardiasis in the metropolis and Tokai regions of Japan ranges from 0.2-0.9% [8,9]. However, the study populations consisted of outpatients and/or inpatients who might not have represented the total population in the area. Furthermore, it is time consuming and expensive to examine thousands of individuals, and it is difficult to obtain consent for stool surveys.

Sykora et al. examined 11 sewage plants at different cities in the USA and detected 642 to 3,375 *Giardia* cysts per liter of samples [10], and Jakubowski et al. used these data to clarify the relationship between cyst number and prevalence in the respective cities [11]. Although these studies are interesting from the viewpoint of simplicity and/or economy, they do not discuss the recovery efficiency of their method or the circadian rhythm of cyst numbers. In this study, we recovered *Giardia* cysts from sewage samples using the immunomagnetic separation method at a fixed time each month and used the results to estimate the prevalence of the intake of *Giardia* infection among inhabitants in Hyogo Prefecture, Japan.

**MATERIALS AND METHODS**

**Sewage Plants**

The present survey was conducted at two different sewage treatment plants (A and B) in Hyogo Prefecture, Japan, during a one-year period (July 2002 to September 2003). Plant A is located on an 44° 10'-m² artificial island...
constructed in 1981 in the southern part of the prefecture. Approximately 15 \( \times 10^3 \) people live on this island, and plant A treats only the sewage from this population. There are dogs and cats but no large animals on the island. On the other hand, plant B is located 22 km west of plant A and treats the sewage of approximately 205 \( \times 10^3 \) people. Many livestock animals such as cows, chickens and pigs are bred in this area. The mean daily flow of plants A and B during the study period was 8 \( \times 10^9 \) and 66 \( \times 10^9 \) L, respectively.

Sample Collection and Concentration

Two hundred ml of sewage was collected from each plant at 10:00 a.m. once a month for one year. Samples were placed in 250-ml conical tubes into which 2 ml of 10\% gelatin solution was added to effectively disperse the substances. The tubes were then capped tightly and their contents mixed thoroughly. In an additional experiment, samples were collected from plant A every two hours for 24 hours to clarify the hourly changes in cyst number. We repeated this experiment twice, on May 28 and August 12, 2003. In addition, sampling was performed for 5 continuous days in plant A to compare cyst number before (April 14-18, 2003) and after the holiday week (May12-16, 2003).

The sewage sample was filtered through a stainless steel sieve with 32 \( \mu \)m pores, then centrifuged at 1,000 \( \times \) g for 10 min. The supernatant was aspirated leaving 4 ml of concentrate in the tube into which 4 ml each of ethyl alcohol and diethyl ether were added (total 12 ml). The samples were then mixed and incubated at 60 \( > \)C for 5 min. After further mixing, the contents were centrifuged at 1,000 \( \times \) g for 10 min, then the pellet was re-suspended in 15 ml of 0.1\% gelatin solution and centrifuged again under the same conditions. Finally, the pellet was suspended in 10 ml of distilled water (Fig. 1).

Separation Procedure and Counting of Cysts

*Giardia* cysts suspended in the 10-ml water sample were purified using an immunomagnetic separation (IMS) method. In brief, samples were incubated with paramagnetic beads (Dynabeads, Dynal, Oslo, Norway) which were coated with antibodies raised against *Giardia*. The resulting complex of cysts and beads was then magnetically isolated and washed. The cysts were then dissociated from the beads, and the beads magnetically removed from the purified sample. The cysts were placed in a well-slide and stained using a fluorescein-labeled monoclonal antibody (EasyStain, Biotechfrontiers, North Ryde, Australia) then observed under a fluorescent microscope (BX50, Olympus, Tokyo, Japan). Round or oval fluorescein isothiocyanate (FITC)-labeled objects measuring 7-10 \( \times \) 12-15 \( \mu \)m with two or more internal characteristic features (2-4 nuclei, median bodies, axonemes) were counted as *Giardia* cysts.

**RESULTS**

The recovery efficiency of this method was calculated by spiking one hundred *Giardia* cysts labeled with red fluorescent dye (Colorseed, Biotechfrontiers, North Ryde, Aus-

![Fig. 1](image-url)
tralia) into 200-ml sewage samples from plant A; this spiking procedure was repeated three times. The average recovery efficiency was calculated as 40% (37, 42, 40, n=3) and the number of cysts per liter of sewage sample was corrected according to this recovery efficiency.

Circadian rhythms in cyst numbers in plants A and B were observed, with a peak at 10:00 a.m. (4,300/L and 7,700/L, respectively) (Fig. 2). The average number of cysts was 1,700/L and 1,000/L, respectively). The ratio of cyst number at 10:00 a.m. to the daily average cyst number was 0.22 and 0.24 respectively, and the mean was 0.23.

Monthly investigation samplings were performed at 10:00 a.m.; therefore, cyst numbers were adjusted to the daily average number. The daily average number of cysts in each examination was equal to the cyst number detected \( \times \) correction factor (0.23; the correction factor is the coefficient used to correct the number of cysts obtained from the 10:00 a.m. sampling period to the daily average).

The seasonal variation in cyst numbers is shown in Fig. 3. The present study revealed that both of the sewage plants studied were positive for *Giardia* cysts, with a mean number of 880/L (range: 370-2,000/L) and 2,000/L (range: 850-3,900/L) in plants A and B, respectively. Cysts were detected throughout the year in both plants, with the number of cysts detected in plant B being higher than that in plant A. Monthly changes in the number of cysts recovered from plants A and B showed two peaks, in one May and the other in December. The number of cysts detected after a holiday week was higher than that before: the average number detected before and after was 1,500/L and 2,500/L, respectively, and the difference was significant (p<0.05 student’s t test) (Fig. 4).

The estimated number of infected individuals on each of the examination days was the daily sewage flow \( \times \) cyst number of each monthly examination / number of cysts excreted by a patient in one day (1 to 2 \( \times \) 10^9; according to Tsuchiya, 1931); the estimated prevalence was the estimated number of infected individuals / population in each plant area.

### Table 1. Estimation of prevalence of giardiasis on examination days in areas covered by plant A and B

<table>
<thead>
<tr>
<th>Plant</th>
<th>Quantity of sewage treated /day (L)*</th>
<th>Number of cysts found (L)**</th>
<th>Population</th>
<th>Estimated number of patients</th>
<th>Estimated prevalence (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>8x10^6</td>
<td>370 (min.) 880 (ave.) 2000 (max.)</td>
<td>15 x 10^3</td>
<td>15-30 35-70 80-160</td>
<td>0.1-0.2 0.2-0.5 0.5-1.1</td>
</tr>
<tr>
<td>B</td>
<td>6x10^6</td>
<td>850 (min.) 2000 (ave.) 3900 (max.)</td>
<td>205 x 10^3</td>
<td>280-560 650-1300 1300-2600</td>
<td>0.1-0.3 0.3-0.6 0.6-1.3</td>
</tr>
</tbody>
</table>

Estimated number of patients=\* \( \times \)**/number of cysts excreted per patient per day (1 to 2 \( \times \) 10^9, according to Tsuchiya, 1931)
Estimated prevalence=estimated number of patients / population in each plant area.
The prevalence of giardiasis in the areas serviced by plants A and B was estimated using the above results (Table 1). Consequently, the average monthly prevalence was 0.1-1.1 and 0.1-1.3%, respectively.

**DISCUSSION**

*Giardia* is widely distributed throughout the world with a varying rate of prevalence [2], and it is one of the most common enteric parasites recovered from humans, especially in tropical areas. However, it has become difficult even in these areas, to obtain consent to sample fecal materials. In the U.S.A., attempts have been made to detect *Giardia* cysts in sewage and to estimate giardiasis prevalence [11], but the sucrose flotation and Lugol’s iodine techniques used are relatively insensitive. For instance, using the sucrose flotation technique, Sykora et al. found highly variable results (0.4 to 77.8%) with a mean of 11.0% [10]. With this in mind, we used a modified immunomagnetic bead method [12] to detect *Giardia* in sewage samples. In the present study, the number of cysts detected after six weeks after the onset of symptoms was 1 $10^7$ with cyst-excreting and non-excreting days occurring alternatively during the course of infection [24]. Using this data as a base, we estimated the mean number of cysts excreted per day per patient. Accordingly, the mean number of cysts excreted for a period of six weeks after the onset of symptoms was $1 \times 10^6$, but when calculations were conducted using only the cyst-excreting days the value was $2 \times 10^6$.

**REFERENCES**


Kimura et al. examined Japanese travelers who visited Nepal and India for more than 10 days and who experienced diarrhea when they arrived back in Japan [16]. Their survey revealed that 69 out of 692 (9%) travelers were positive for *Giardia* cysts. *Giardia* is the most common enteric protozoan parasite in Nepal and India [17,18,19,20] and has remained top of the list for the past decade [17]. Similar imports of *Giardia* infections into developed countries have been reported elsewhere [3]. The reported *Giardia* cyst shedding rates in calves ranges from 80 [21] to 100% [22] in British Columbia, Canada. However, since the area serviced by plant A is an isolated artificial island where livestock are not bred, we can rule out the possibility of sewage contamination by such animals. The hourly variation results obtained in this study also clearly demonstrated that cyst concentrations are related to human activity, and the findings strongly suggest that the *Giardia* cysts were excreted from inhabitants of this island. This result agrees with the earlier study of Gassmann and Schwartzbrod who suggested that a peak of *Giardia* cysts was observed at 10:00 a.m. in Nancy, France [23].

It is difficult to assess the number of cysts excreted daily from observations of a single infected individual. Therefore, we calculated the number of infected individuals based on the recovery efficiency, monthly survey, and variation in cyst numbers at different sampling times. Tsuchiya calculated that the number of cysts excreted from an individual/day varied from 0-7 $10^7$ with cyst-excreting and non-excreting days occurring alternatively during the course of infection [24]. Using this data as a base, we estimated the mean number of cysts excreted per day per patient. Accordingly, the mean number of cysts excreted for a period of six weeks after the onset of symptoms was $1 \times 10^6$, but when calculations were conducted using only the cyst-excreting days the value was $2 \times 10^6$.

The infection rate of *Giardia* in the areas serviced by plants A and B was estimated as 0.1-1.1 and 0.1-1.3%, respectively, and these results were consistent with the results obtained from fecal examinations in Japan (0.2-0.9%) [8,9]. This study revealed that sewage samples can be used to estimate giardiasis in a particular service area (community) using the IMS method. The method developed for this survey was easy, cost-effective and time saving, and it allowed for accurate estimations of the prevalence of giardiasis. We believe it can be used for future surveys of the prevalence of giardiasis in other areas.

The prevalence of giardiasis in the areas serviced by plants A and B was estimated using the above results (Table 1). Consequently, the average monthly prevalence was 0.1-1.1 and 0.1-1.3%, respectively.


